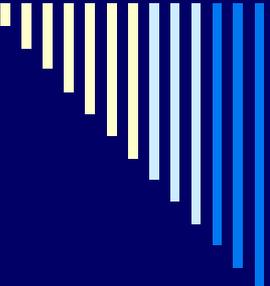


IEF Electrophoresis

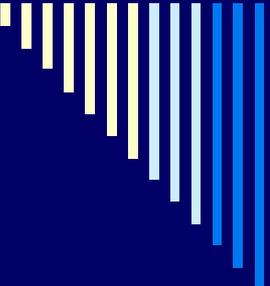
2016 SCST

Genetic Testing Super Workshop



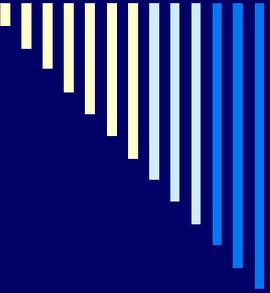
Isoelectric Focusing (IEF)

- Can use polyacrylamide or agarose gels
- Based on molecules that can be positively or negatively charged (amphoteric)
- For example: proteins, enzymes, peptides
- The net charge of protein is the sum of the positive and negative charges of the amino acid side chains.
- When the net charge of the protein is 0 that is the proteins isoelectric point.
- When a protein has reached its isoelectric point, it will stop migrating through the gel at a certain pH point.



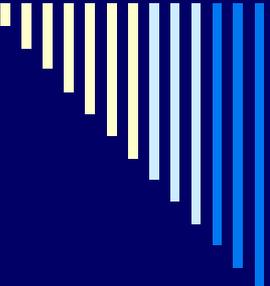
How does the pH gradient work in the gel matrix

- Carrier ampholytes are in the gel matrix that are low molecular weight and have closely related isoelectric points
- When electricity is applied to the gel the ampholytes forms a pH gradient in the gel.
- When an amphoteric protein from a sample is no longer charged the electrical current will not have an effect on it. If that protein were to diffuse back away from its pI point, it will gain a net charge again, and go back to that same pI and pH point in the gel. Thus, the term FOCUSING.



Four processes are involved:

- Extraction
- Focusing
- Staining
- Analysis



Extraction Process

- Weigh 10 seeds of sample to determine extraction amount used
- Load a single seed into each well of a 48 well plate
- Crush seed
- Extract plates
- Vortex and let incubate overnight

Supplies needed for extraction process



Crushing seed

Manual seed crusher



Manual seed cutter

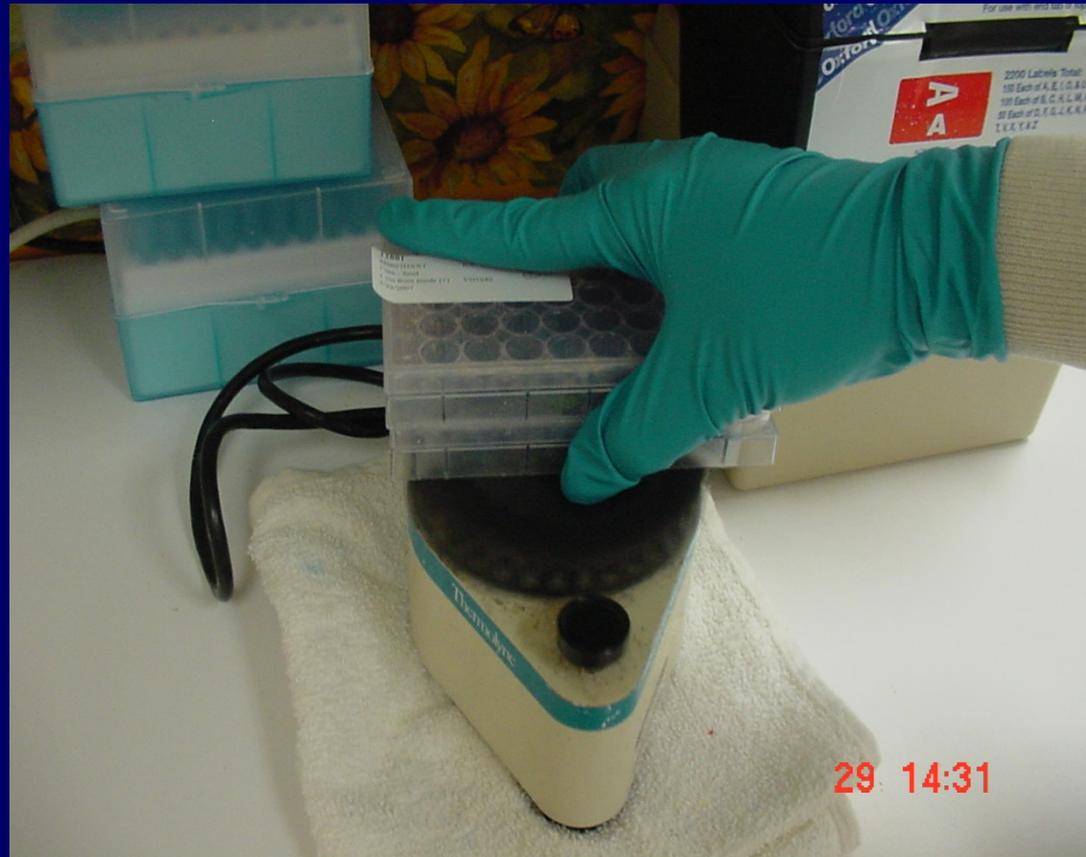


23 15:30

Crushed corn seed

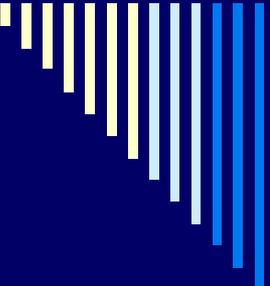


Vortexing plates after adding extraction solution



Supplies for making control samples

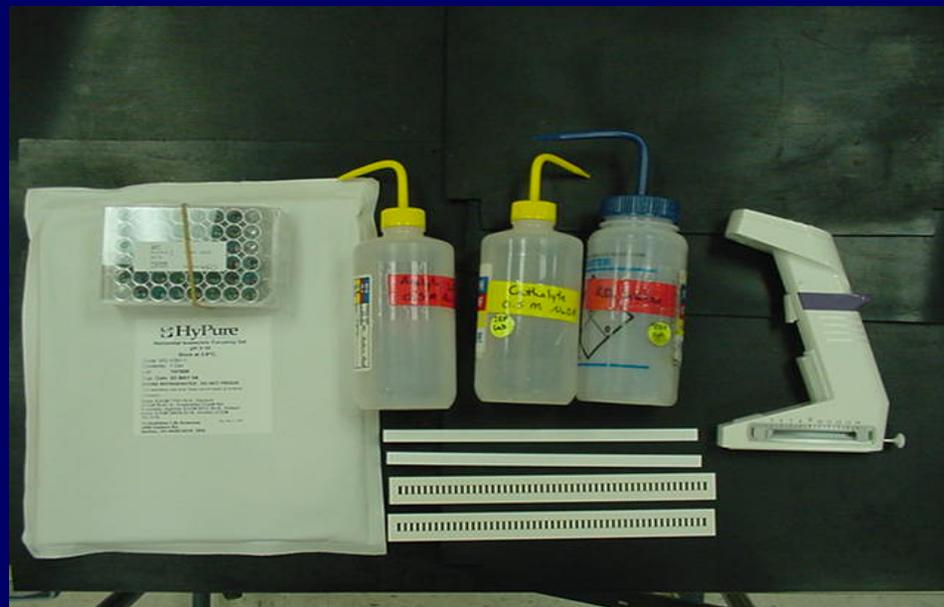




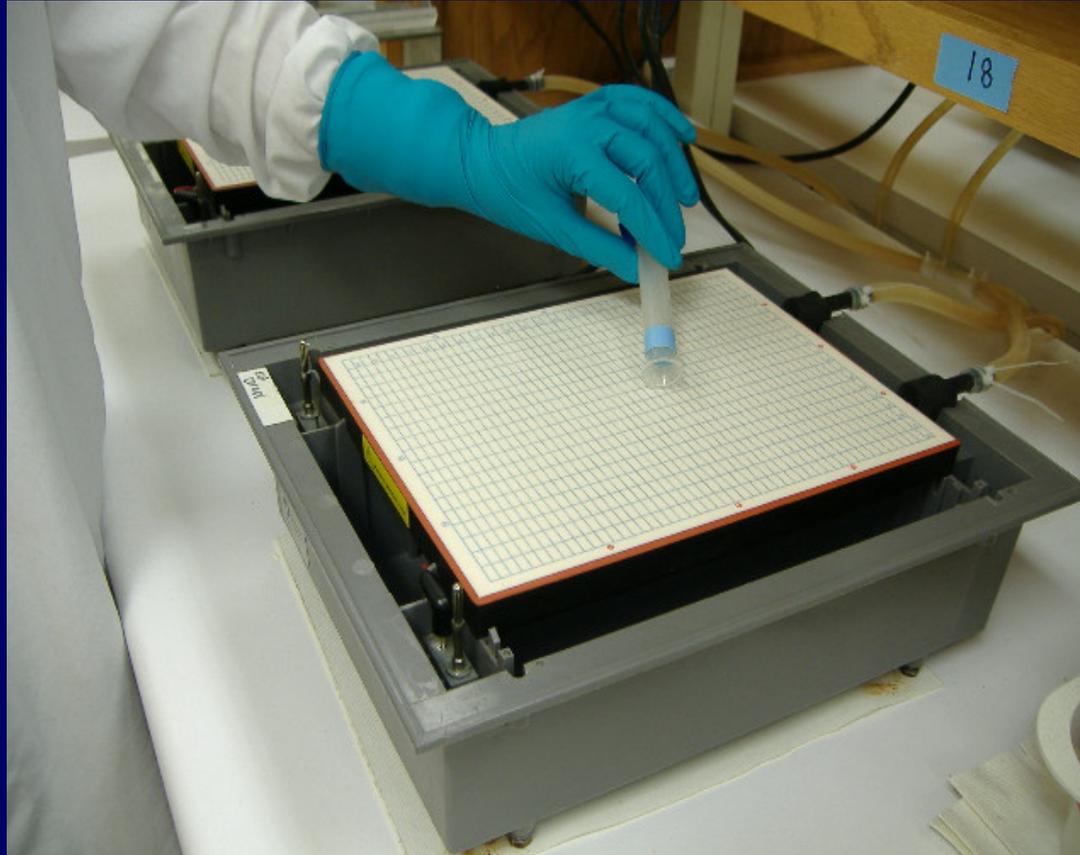
Focusing

- Label gel with date, sample #, and gel type
- Make sure data sheet matches the plates
- Load the gel
- Run the gel
- Place in TCA fixative solution
- Place in 2 water rinses

Supplies for focusing gels



Pour ~2 ml water on plate

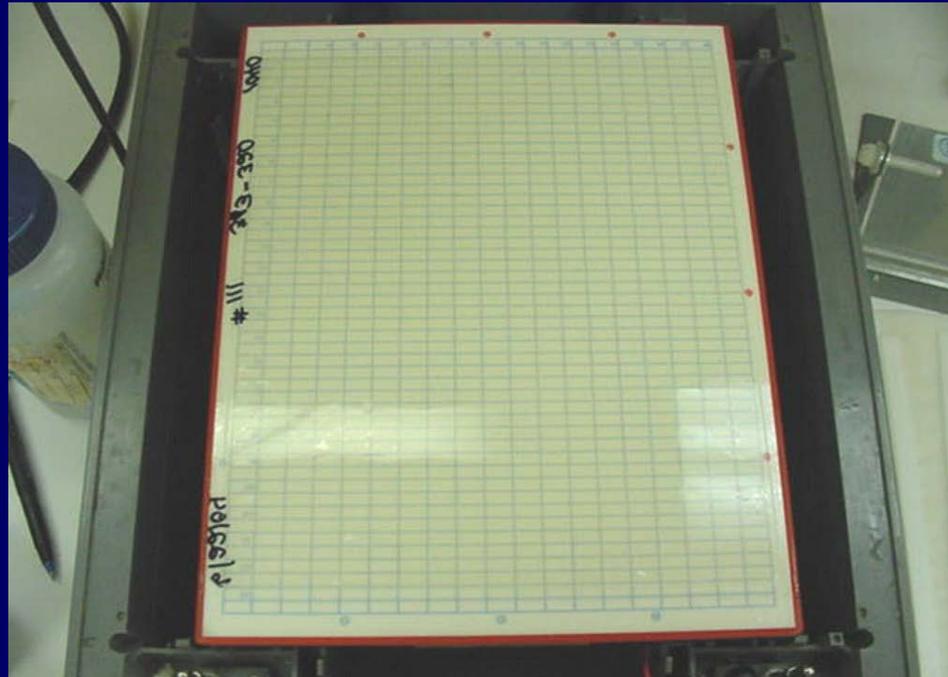


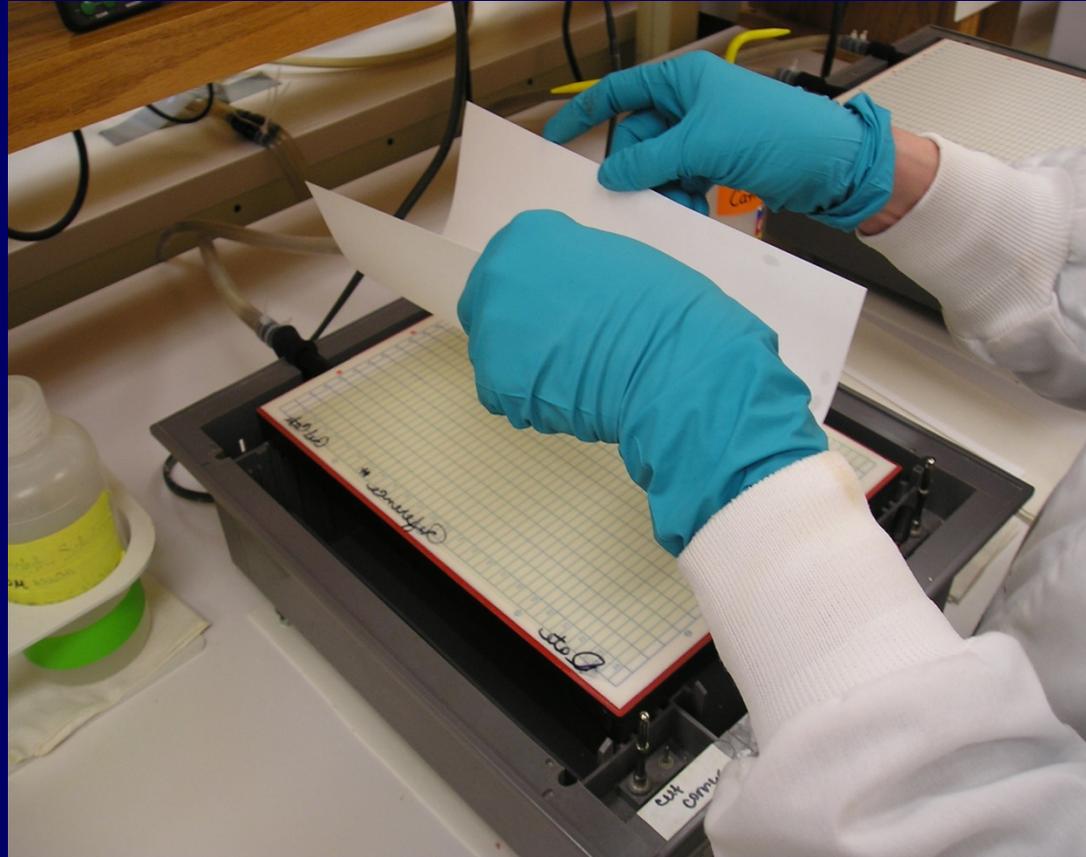
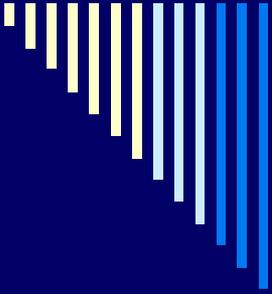
Circulating water bath



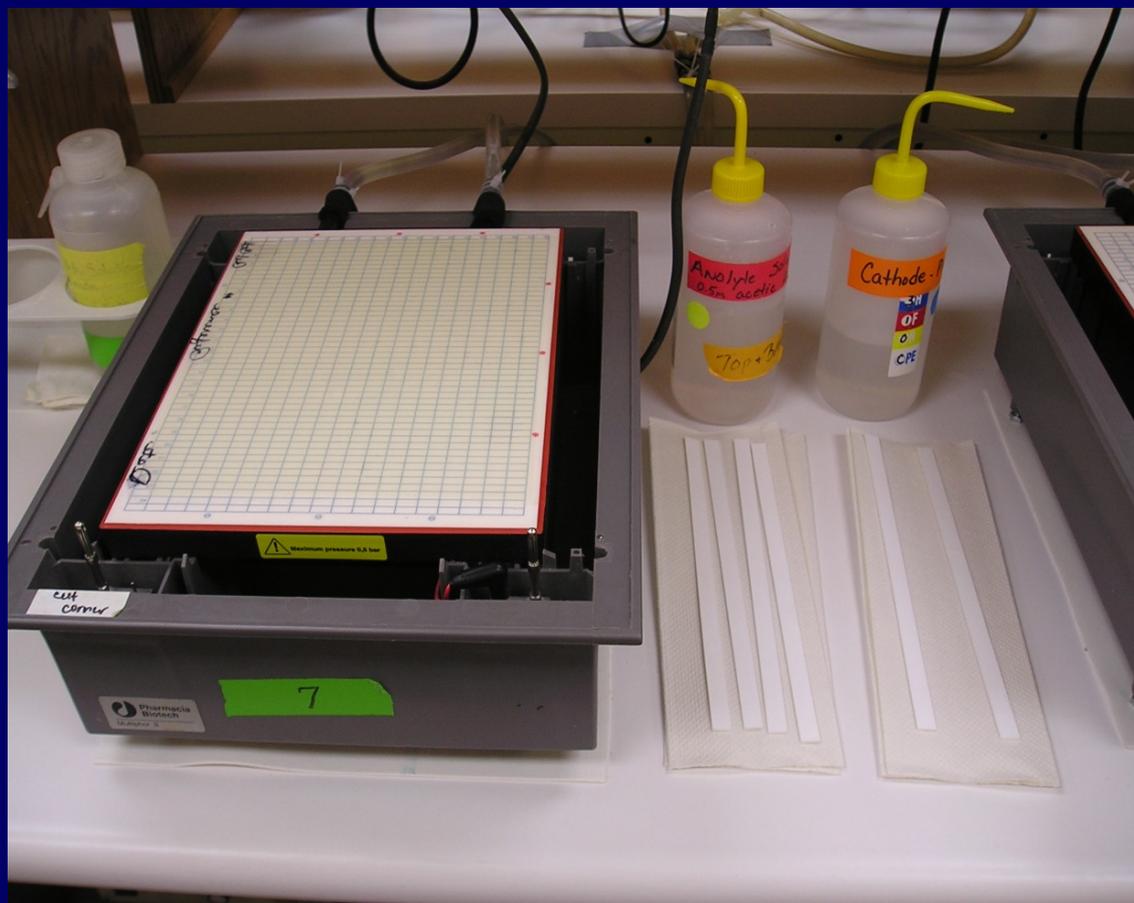
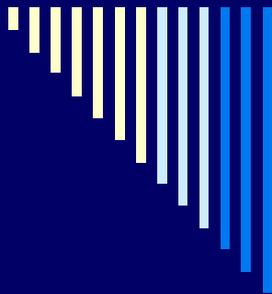


Place gel on multiphor using grid pattern on plate









Soaking wicks with anode and cathode solutions

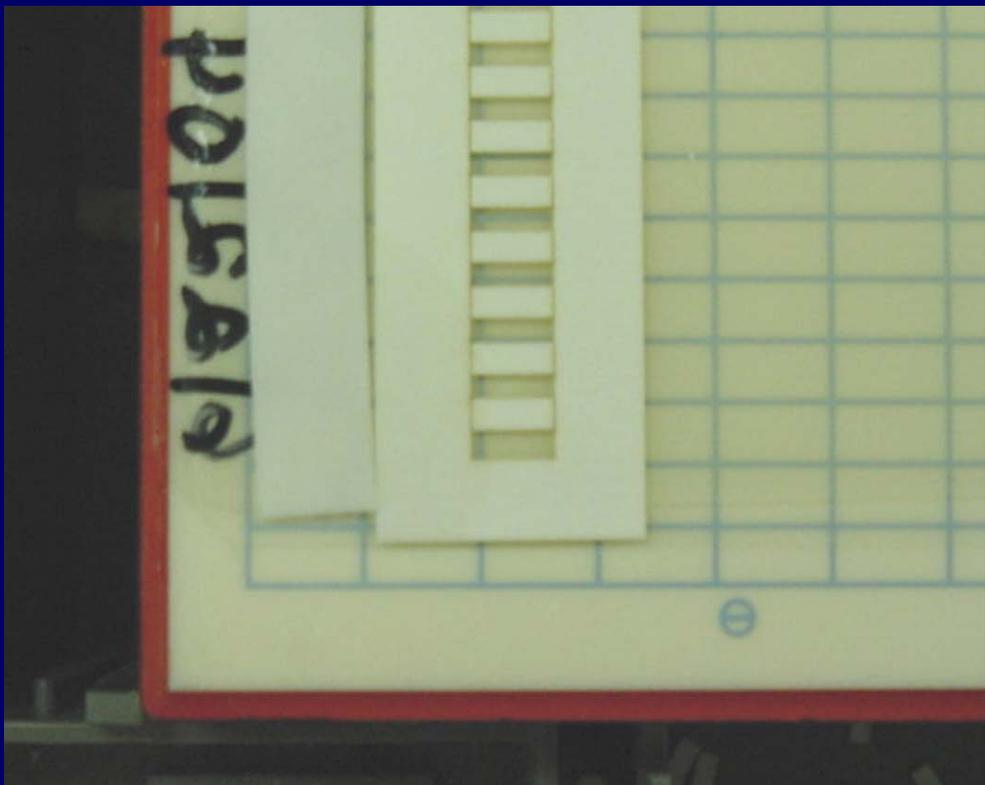
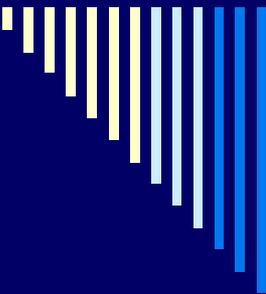


Placing wicks on gel

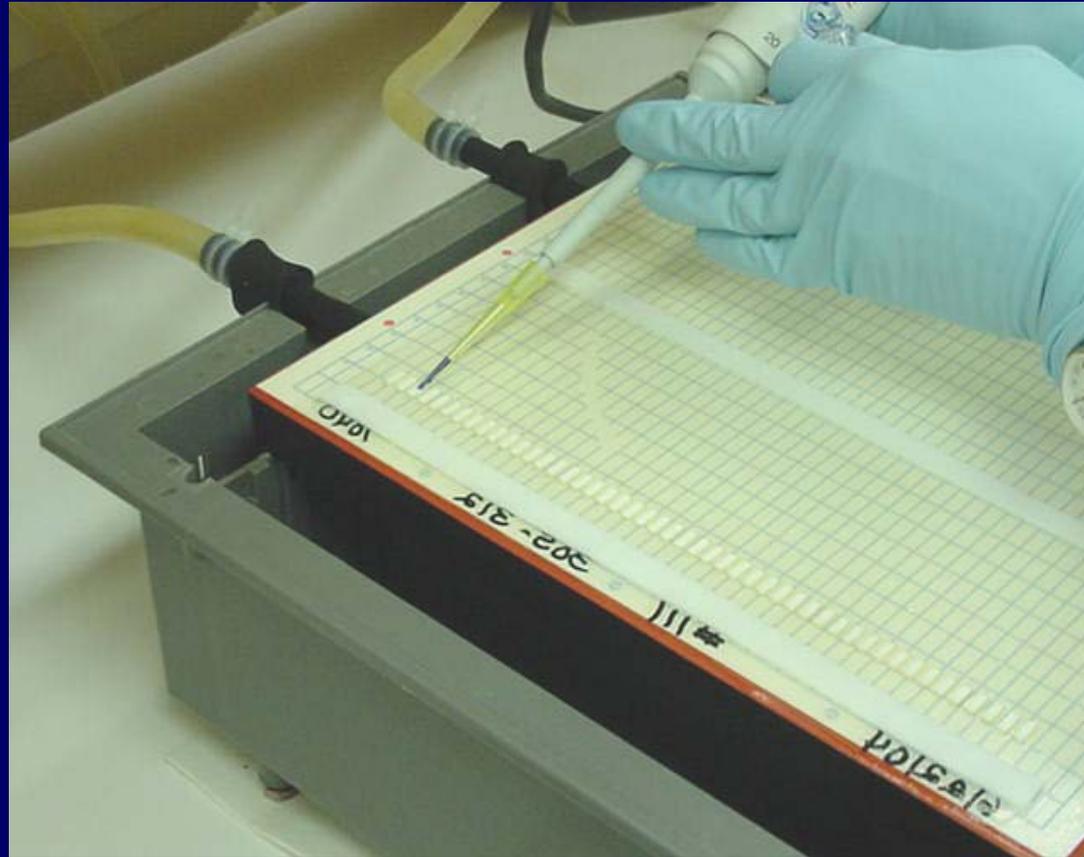


Placing template on gel

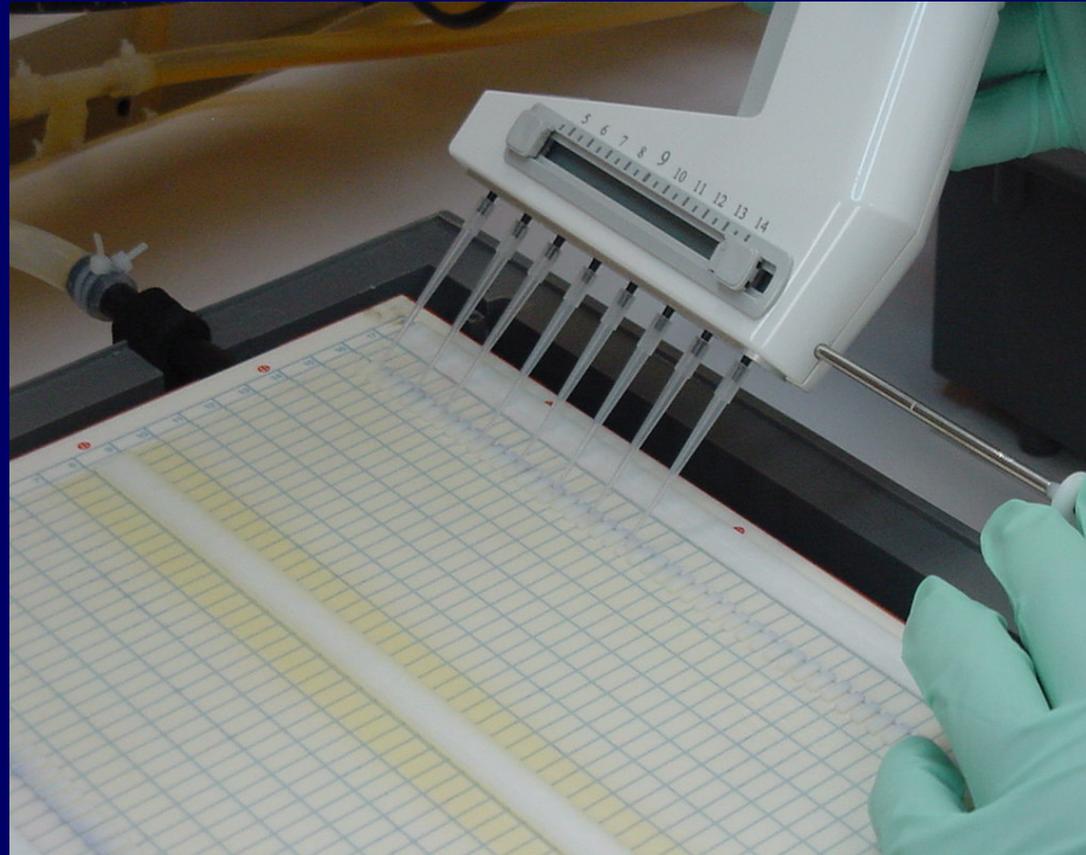




Pipetting sample onto template



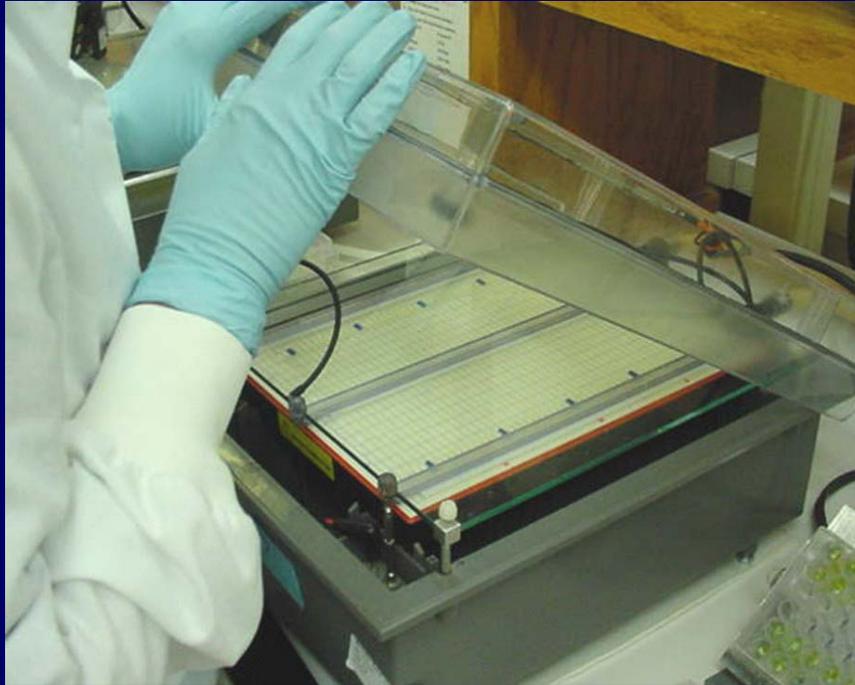
Pipetting sample onto template using multi-channel pipette.



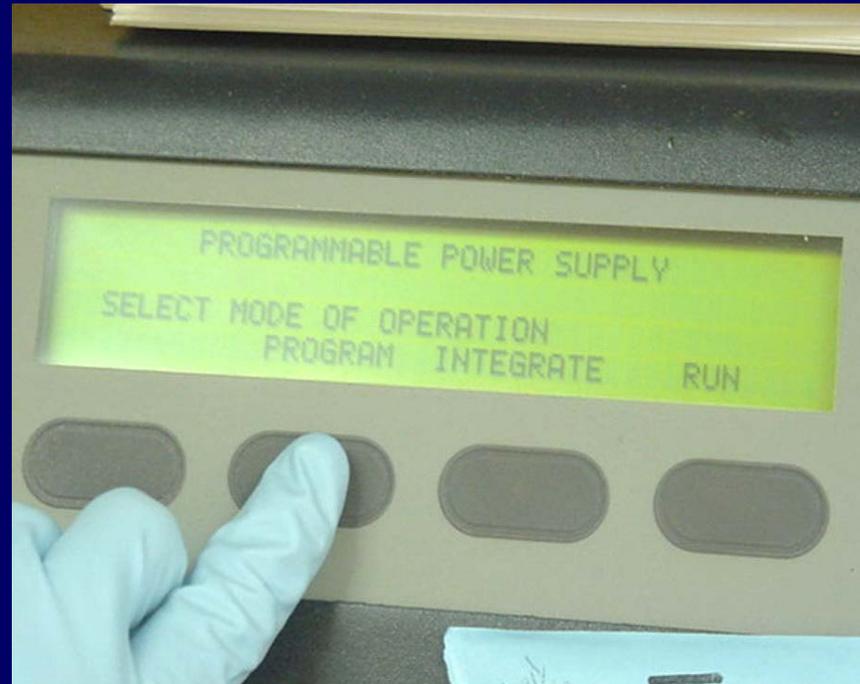
Placing anode and cathode wires onto gel.



Putting lid on which completes the circuit



Starting up the power supply



Record readings on data sheet

Date Sample Run _____
 Gel Lot Number _____
 Gel Expiration Date _____
 Multiphor Number _____
 Power Supply Letter _____
 Amount Sample/Well _____
 Initials _____
 Comments _____

Gel Run Readings				Gel Run Readings			
45 min	Time to Run		45 min	Time to Run			
Start	Stop		Start	Stop			
100	Watts	100	Watts		Watts		
48.9	Volts	10.26	Volts		Volts		
20.7	mAmps	9.9	mAmps		mAmps		
11	Temp	10	Temp		Temp		
10:35	Time	11:20	Time		Time		

Stain Conditions
 Stain Method _____
 Date Sample Stained _____
 Initials _____

Date Sample Analyzed _____
 Initials _____
 Comments _____

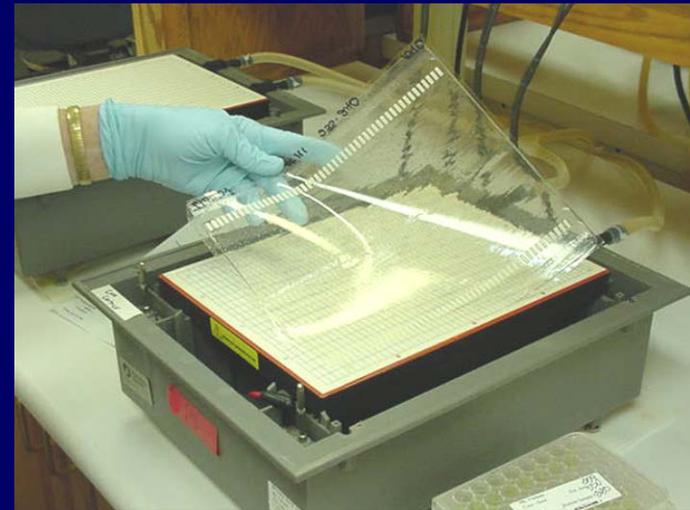
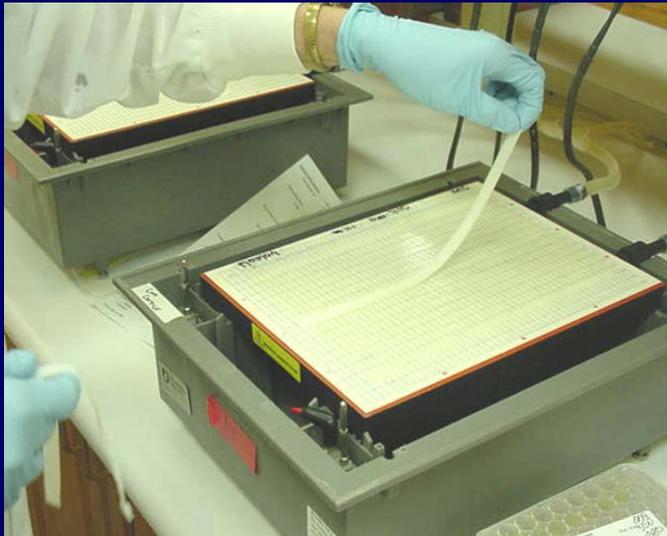
Gels running



Blotting the gel. (with towel or blotting paper)



Taking gel off once it is done with the run



Gels rocking in TCA fixative step.



TCA fume hood

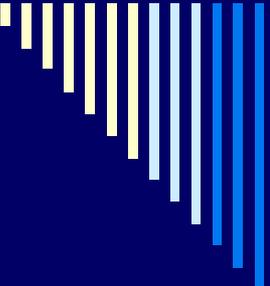


Gels rocking in water rinses



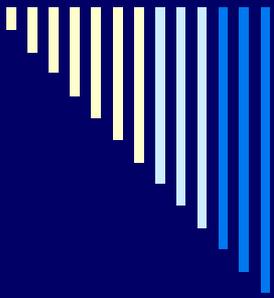
Once gels are out of water rinses, they hang dry overnight or go into a dryer





Staining Process

- Make all solutions for staining
- Place gels in dishes and heat
- Start staining
- When gel is stained, place in an acetic acid stop solution
- Rinse gel with water
- Blot dry



Sodium Carbonate solution



Chemical prep for staining



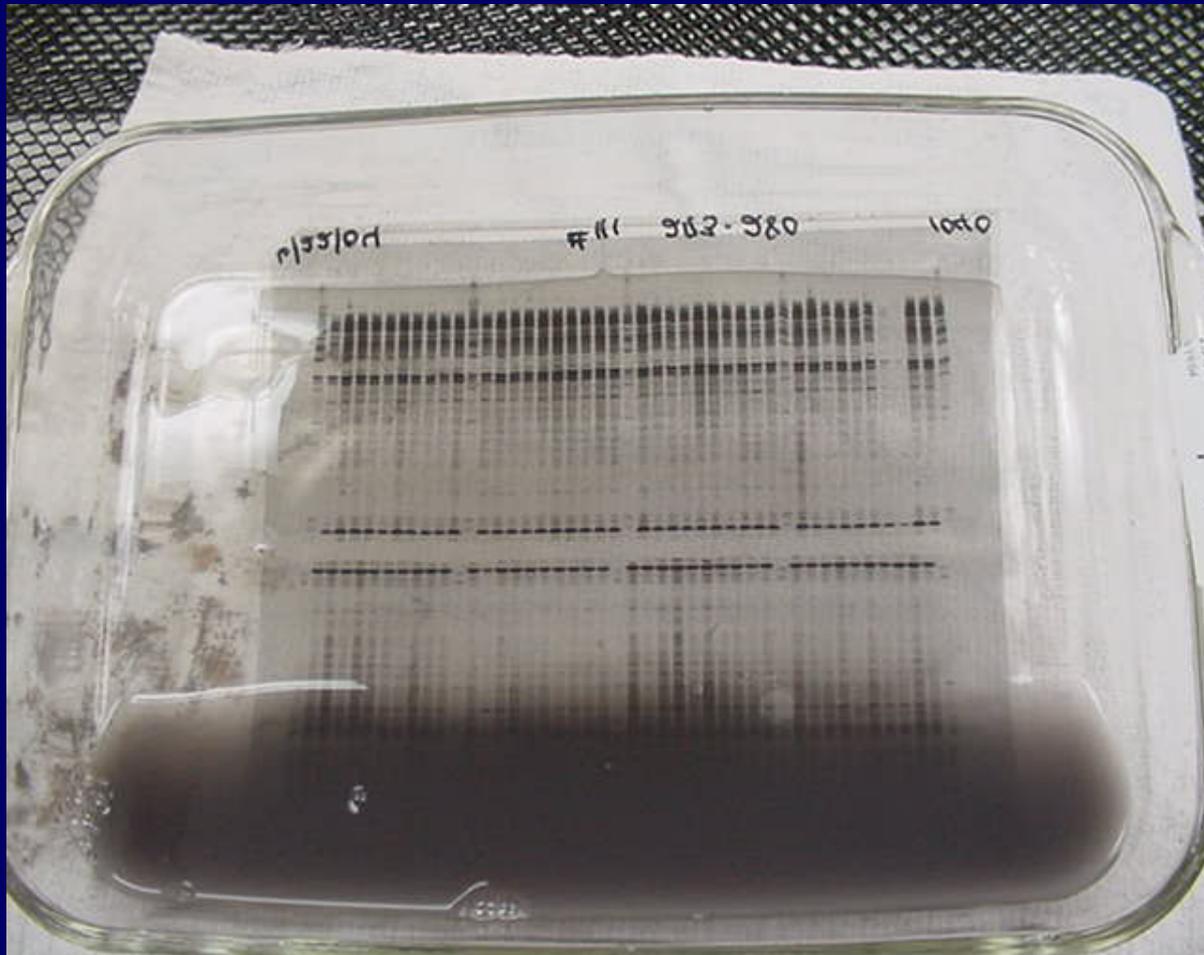
Staining hood



Pouring staining solution onto gel



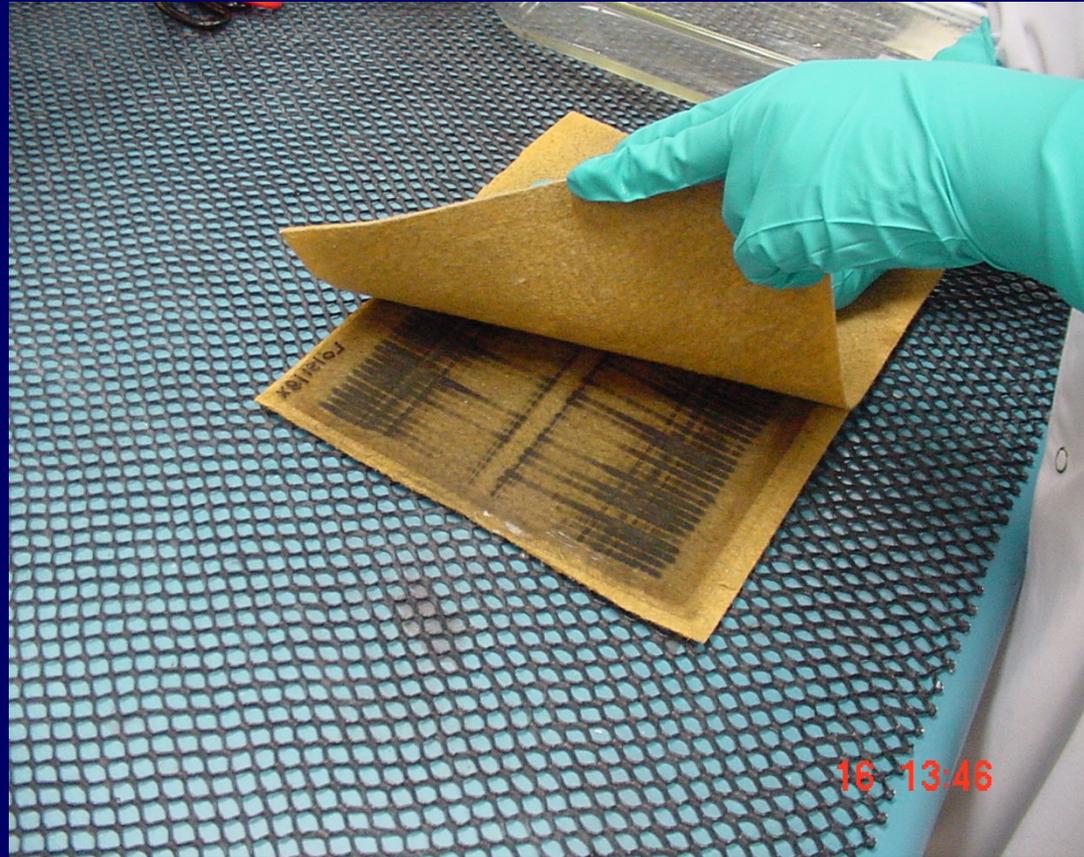
Stained Gel



Pouring stop solution in waste container

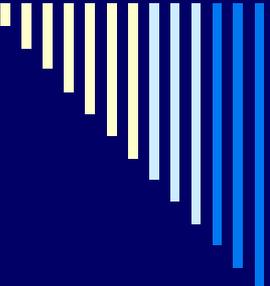


Blotting gel after stop solution step



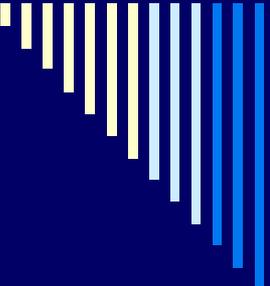
Gels that are ready for analysis





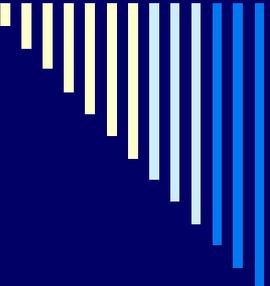
What can we see with IEF gels

- Can determine female selfing
- Can determine if a parent line is fixed or not (segregation present)
- Can determine off types
- Can have results within 24 hours (can crush and extract sample same day received and focus/stain/analyze the next day)



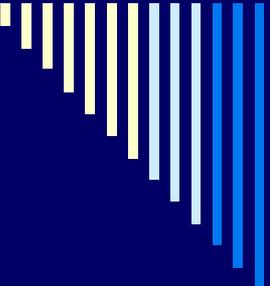
What can we see (cont.)

- Can compare foundation Gen0 against Gen1 and Gen2
- IEF gels will keep indefinitely. This is valuable for use as reference material.
- We have every gel that has been run on every material that has been tested since using IEF for genetic purity testing.



Nothing ever goes wrong!

- ❑ If you switch the anode and cathode solutions when loading your gels, the readings will be off. (Volts go down, amps go up)
- ❑ If you forget to put a gel in TCA soln you will have a practically blank gel when staining
- ❑ If a platinum wire breaks, an anode/cathode isn't plugged in right, or a gel is taken off too soon-the gel will not run the whole way
- ❑ A gel can start sparking
- ❑ Power supply is set for the wrong wattage



Safety considerations

- Formaldehyde-regulated chemical
- TCA-pH 1
- We wear cuffed lab coats and long cuffed gloves

Questions? Comments?